



STUDY OF PATHOGENIC MICROBES FROM EDIBLE OYSTER (CRASSOSTREAGRYPHOIDES) OF KELVA COAST;PALGHAR, MAHARASHTRA

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Abstract

Present study on edible oysters (*Crassostreagryhoides*) was conducted at kelva coast, Palghar, Maharashtra. 12 samples were processed from 2016 to 2018 for bacterial study. A total of 44 isolates were isolated. The occurrence of *Aeromonas* spp. was (29%) followed by *Pseudomonas* spp. (25%), *Enterobacterkobei* (23%), *Providenciavermicola* (14%) and *Bacillus flexus* (09%). In the present study an attempt was made to enumerate the bacteria along with health hazard if any.

Key words: Pathogenic microbes, Kelva, Health hazards.

Introduction

The edible oysters are very popular as raw and processed food in the South Indian states, particularly in Goa, in the South East Asian countries, Europe, Australia and USA. Naturally, oyster accumulates microorganisms during the process of filter feeding. These shellfish are prone to harsh environmental contamination by fecal pathogens like *Salmonella* sp. *Shigella* sp. and *Escherichia coli* (Musa et al., 2008). Illnesses due to food-borne contamination frequently occur, but rarely documented. *E. coli* and *Salmonella* sp. could extensively spread inside the body of human beings consuming oyster and lead to serious infection to the human and death (Forsythe, 2002). On the other hand, *Vibrio* sp. may be transmitted to humans by the ingestion of raw seafood. Oysters from these waters are often incriminated in human diseases since the oysters are commonly consumed as raw (Haldy, 1997).

Farely good amount of work has been carried out on edible oyster from different developed countries and rarely from India and not from Palghar district of Maharashtra. Since edible oyster are cheaply available sea food, it is becoming the good source of food to the people in the vicinity specially to the poor. It is intended in this paper to enumerate the pathogenic microbes from the edible oyster *Crassostreagryhoides* and in turn the health hazards if any.

Study Area

This study was carried out at KelvaCoast, Palghar.Kelva is the tourist place of PalgharTaluka which is situated at the coastal area of Arabian sea having geographic coordinates of 19°37'9"N 72°43'23"E. Figure No. 1 and 2.

Figure.1 and 2:- Kelva coast of PalgharTaluka.



Materials and methods

A total of 12 samples of edible oysters (Crassostereagryphoids) were collected from kelvacoast during the period from 2016-18. Samples were collected from the rocks with the help of hammer and chisel, packed in polythene zip wrap bag,labelled, kept in the ice box and immediately transported to the Zoology department research Laboratory of S.D.S.M. college, Palghar within an hour. Further samples were processed for the Total Plate count and differential pathogens with standard protocol. The 10 gram of edible oyster already depurated from shell was transferred to a sterile beaker to which 90 ml of normal saline solution (NSS) was added. The samples were serially diluted by 10 fold serial dilution method in the normal saline solution up to 10^{-7} . The 10^{-7} dilution were used in 0.1 ml quantities for the Standard Plate count (SPC) on Plate count agar (PCA). The agar plates were inoculated by pour plate method and incubated at 37°C for 24 hrs. The 10^{-4} dilutions were taken for plating following differential media simultaneously during processing of the samples, Baird Parker agar, Slanetz and Bartley agar, Macconkey agar, Violet red bile agar andThiosulphate Citrate Bile Sucrose agar.Salmonella Shigella agar and Xylose lysine deoxycholate agar was streaked after enriching the sample in selenite cystine broth at 37°C for 18 hrs. Suspected microbes were further identified by growth pattern and morphology of the colony, direct microscopic examination and different biochemical tests (Cowan and Steel.,1993 and Hi-media Product information 1992, 2016, 2018).Authentication of representative organism was carried out at geneOmbio lab Baner, Pune.

Result

Total 12 samples of edible oyster(Crassostreagryphoides) were collected from Kelvacoast and the same were processed for Total viable count(T.V.C) and



different pathogenic bacteria. The T.V.C ranged from 1×10^7 to 35×10^7 cfu/ml. Out of 12 samples processed, the total 44 isolates were isolated. *Enterobacter* spp. and *Pseudomonas* spp. Occurred in maximum number of samples (10 each) followed by *Aeromonas* spp. (7), *Providencia* spp. (5) and *Bacillus* spp. (4). The sample wise number is shown in Table 1 and % wise pathogenic microbes in Table 2 and figure 3.

Gram Positive Rods

From the total 44 isolates isolated from the 12 edible oyster samples *Bacillus flexus* were encountered in 04 samples which constituted 09% of the total organisms.

Gram Negative Rods

From the total 44 isolates isolated from the 12 edible oysters samples *Aeromonas* spp. encountered in maximum number and constituted (29%) followed by *Pseudomonas* spp. (25%), *Enterobacter kobei* (23%) and *Providencia vermicola* (14%).

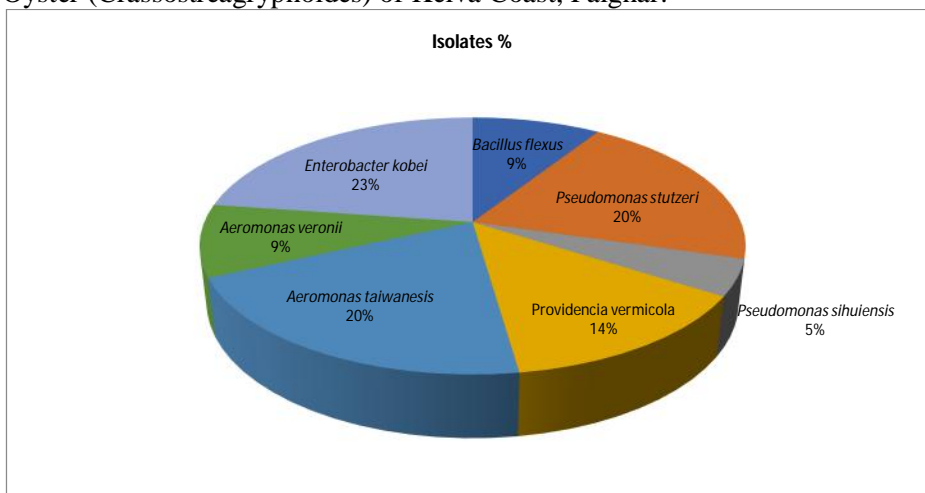
Table 1. Sample wise microbes of edible oyster (*Crassostrea* spp.) from Kelva coast of Palghar

Sample No.	Name of microbes	Total No. of microbes	T.V.C= $\times 10^7$ (cfu/ml)
1	<i>Pseudomonas stutzeri</i> , <i>Providencia vermicola</i> , <i>Enterobacter kobei</i>	03	10
2	<i>Pseudomonas stutzeri</i> , <i>Pseudomonas sihuensis</i> , <i>Providencia vermicola</i> , <i>Aeromonas veronii</i> , <i>Aeromonas taiwanensis</i> (2), <i>Enterobacter kobei</i>	07	20
3	<i>Pseudomonas stutzeri</i> , <i>Pseudomonas sihuensis</i> , <i>Bacillus flexus</i> , <i>Aeromonas taiwanensis</i> , <i>Enterobacter kobei</i>	05	20
4	<i>Pseudomonas stutzeri</i> , <i>Providencia vermicola</i> , <i>Bacillus flexus</i> , <i>Aeromonas veronii</i> , <i>Aeromonas taiwanensis</i> , <i>Enterobacter kobei</i>	06	25
5	<i>Providencia vermicola</i> , <i>Aeromonas taiwanensis</i> , <i>Enterobacter kobei</i>	03	30
6	<i>Providencia vermicola</i> , <i>Bacillus flexus</i>	02	08
7	<i>Pseudomonas stutzeri</i> , <i>Aeromonas veronii</i> , <i>Aeromonas taiwanensis</i> , <i>Enterobacter kobei</i>	04	35
8	<i>Pseudomonas stutzeri</i> , <i>Aeromonas taiwanensis</i> , <i>Enterobacter kobei</i>	03	06
9	<i>Pseudomonas stutzeri</i> , <i>Enterobacter kobei</i>	02	05
10	<i>Providencia vermicola</i> , <i>Enterobacter kobei</i>	02	16
11	<i>Pseudomonas stutzeri</i> , <i>Enterobacter kobei</i>	02	01
12	<i>Pseudomonas stutzeri</i> , <i>Bacillus flexus</i> , <i>Aeromonas taiwanensis</i> (2), <i>Aeromonas veronii</i>	05	12
	TOTAL	44	

Table 2. Percentage of different pathogenic microbes isolated from Edible Oyster (*Crassostrea gryphoides*) of Kelva Coast, Palghar.

Sr. No.	Name of Isolates	No. of isolates	%
1.	<i>Bacillus flexus</i>	04	09
2.	<i>Pseudomonas sihuiensis</i>	02	05
3.	<i>Pseudomonas stutzeri</i>	09	20
4.	<i>Aeromonas veronii</i>	04	09
5.	<i>Aeromonas taiwanensis</i>	09	20
6.	<i>Providencia vermicola</i>	06	14
7.	<i>Enterobacter kobei</i>	10	23
	G.T.	44	100

Figure 3. Diagrammatic percentage of different microbes isolated from Edible Oyster (*Crassostrea gryphoides*) of Kelva Coast, Palghar.



Discussion

Gram Positive rods

From the total 44 isolates isolated from 12 samples of edible oyster only 04 isolate were of gram positive rod *Bacillus flexus*, which constituted 09% of the isolates. *Bacillus flexus* is an aerobic, Gram-variable, rod-shaped, endosporeforming, oxidase positive bacteria. The endospores are ellipsoidal, located in central/paracentral, unswollen sporangia. These bacteria may be isolated from feces (poultry) and soil (Wikipedia).

Gram Negative rods

Gram negative rods constituted more than 90% of the total isolates. The maximum number shared by *Aeromonas* spp. (29%) Members of the



genus *Aeromonas* have been associated with a wide spectrum of enteric and non-enteric diseases, in both immunocompromised and immunocompetent patients (Janda and Abbott, 1996, 1998). *Aeromonas veronii* was originally described by Hickman-Brenner et al. (1987) as a novel species in the genus *Aeromonas* that had previously been referred to by the Centers for Disease Control as Enteric Group 77.A. *veronii* biovar *veronii* has rarely been isolated from humans and little information is available regarding its occurrence in clinical specimens and its ability to cause infections. In the original study (Hickman-Brenner et al., 1987), it was isolated from various non-sterile sites, such as wounds, faeces, sputum, maxillary sinus and endotracheal tube, in which its clinical significance remains unclear. In recent years, numerous cases of *Aeromonas* infection have been described in Taiwan (Huang et al., 2007; Wu et al., 2007). (Figueras et al., 2000) initiated the investigation to identify genetically a group of extra intestinal *Aeromonas* strains isolated in the National Cheng Kung University Hospital (Tainan, Taiwan) using a previously described 16S rRNA gene RFLP method *Aeromonas taiwanensis* was first described in 2010 on the basis of one strain (A2-50^T = LMG 24683^T = CECT 7403^T) recovered from the wound of a hospitalized Taiwanese patient (Alperi et al., 2010). So far, one additional clinical fecal and four environmental *A. taiwanensis* isolates have been recorded (Senderovich et al., 2012 and Beaz-Hidalgo et al., 2012).

11 isolates of *Pseudomonas* spp. were recorded from 12 samples of edible oysters. Out of 11 isolates 9 isolates were *Pseudomonas stutzeri* and 2 were of *Pseudomonas sihuiensis* which constituted 20% and 5% respectively. A strain of *Pseudomonas stutzeri* was isolated from the leg ulcers (Lapaget et al., 1968). *Pseudomonas stutzeri* is a Gram-negative, rod-shaped, motile, single polar flagellated, soil bacterium first isolated from human spinal fluid. (Lehmann et al., 1896 and Sijderius et al., 1946) It is a denitrifying bacterium, (Lalucat et al., 2006) and strain KC of *P. stutzeri* may be used for bioremediation as it is able to degrade carbon tetrachloride (Sepulveda-Torres et al. 1999).

Pseudomonas sihuiensis is novel spp. Isolated from a forest soil in sihui city, South China (Wu, Min et al., 2014). No further clinical report is reported of *Pseudomonas sihuiensis* and clear function of this spp. In the present study it is being reported and might be first ever finding in edible oyster from Kelva coast of Maharashtra, India.

10 isolates of *Enterobacter kobei* were recorded in the present finding which is 23% of the total isolates. Harald Hoffmann et al., 2005 first time reported patient with a history of urinary bladder operation developed a urosepsis with an *Enterobacter* isolate displaying the *E. cloacae* phenotype.

Providencia vermicolaris recorded six times and constituted 14% of the total isolates isolated. *Providencia* is a genus within the *Enterobacteriaceae* family closely



related to the Providencia and Morganella genera and later from a diseased rohu fish (Muderet al.,1991), but this species has not been implicated in human infection and have not been reported to cause human infection (<http://www.antimicrobe.org/b227.asp>).

Conclusion

This piece of study is the first attempt to highlight the occurrence of pathogenic microbes from the edible oysters from kelvacoast of Palghar which in turn might be affecting the health of the sellers/consumers. Further in depth study is required on the edible oysters to link to the health hazards.

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